

THE EFFECTIVENESS OF SOME MEDICINAL PLANTS ON BODY PERFORMANCE, HEMATOLOGICAL, ILEUM MORPHOLOGY AND IMMUNE STATUE OF JAPANESE QUAIL

Mahbuba A. M¹A. Ab. Jabbar²N. A. Mustafa³

Prof

Lecturer

Assist. Prof.

^{1,3} Dept. of Animal Res., Coll. of Agri. Engin. Scie., Salahaddin University-Erbil/ Iraq² Dept. of Med. Lab. Tech., Erbil Tech. Health Coll., Erbil Polytech. University-Erbil/ Iraq¹Mahbuba.mustafa@su.edu.krd,²Ahmed.abuljabbar @epu.edu.iq³nidhal.mustafa@su.edu.krd

ABSTRACT

This study was aimed to investigate and compare the effect of medicinal plants extract: Hero flowers (*Alcea kurdica*) in the north of Iraq, Thyme (*Thymus bovie*), and Bay leaf (*Laurus nobilis*) extract with two antibiotic drugs Rifaximine and Ranitidine in quail's drinking water by emphasizing the productive performance, and physiological status of the quails at grower and developer stages till 10th weeks of age, a total of 90 quails were distributed into six treatments, each treatment with three replicates of 15 chicks in each. Data analysis showed significantly increased body weight gain (237.5, 224.1, and 234.9) g for Hero, thyme, and Bay leaf, respectively. In comparison with the control (188.3) g. quail's drunk plant extract mixed water had better hematological and the ileum profiling (goblet cells numbers, Villus height, and Crypt depth) and significantly higher serum concentrations of total protein, globulin, and antibodies titers against some bird virus diseases in comparison with positive control. Also, significantly lower level of albumin, ileum content of *E. Coli*, and Enterococcus count in plant treated quails in comparison with the control T0.

Key words: medicinal herbs, body performance, blood traits, ileum histology, body weight.

محبوبة وأخرون

مجلة العلوم الزراعية العراقية- 2022: 53(4):724-731

فعالية بعض النباتات الطبية على أداء الجسم، وحالة الدم، التشريح النسيجي للفانفي و المناعة في السمان الياباني

³ نضال عبدالغني مصطفى² احمد عبدالجلال عبد الجبار¹ محبوبة عبدالغني مصطفى

أستاذ مساعد

مدرس

أستاذ

^{1,3} قسم الثروة الحيوانية، كلية علوم الهندسة الزراعية، جامعة صلاح الدين - أربيل² قسم تقنيات المختبرات الطبية، كلية أربيل للتقنيات الصحية، جامعة بوليتكنيك أربيل، أربيل، 44001، العراق

المستخلص

هدفت هذه الدراسة لمقارنته تأثير مستخلص النباتات الطبية: زهرة الخطمية أو (هيرو) في شمال العراق (*Alcea kurdica*)، الزعتر *Thymus bovie*، مستخلص أوراق الغار *laurus nobilis* مع مضادين حيويين Rifaximine و Ranitidine في ماء شرب طائر السمان من خلال تأثيرهم على الأداء الإنتاجي، والحالة الفسلجية لطائر السمان في مرحلتى النمو والتطور لغاية عشرة أسابيع، تم توزيع 90 طائر سمان الى ستة معاملات، كل معاملة احتوت على ثلاث مكررات، بمعدل 15 طائر في كل معاملة. أظهر تحليل البيانات زيادة معنوية في زيادة وزن الجسم (237.5 و 224.1 و 234.9)غم لزهرة Hero و الزعتر وأوراق الغار بالنتائج مقارنة بمعاملة السيطرة (188.3) غم. شرب طائر السمان لمستخلصات النباتات في الماء كان له خصائص دم أفضل وخصائص التقطيع النسيجي (عدد خلايا الكأسية، ارتفاع الزغابة، وعمق خبايا ليبركن)، و ارتفاع تراكيز مصد الدم من البروتين الكلي والجلوبيولين والأجسام المضادة ضد بعض أمراض فيروس الطيور مقارنة بمعاملة السيطرة. في حين وجد انخفاض ملحوظ في مستوى الألبومين، ومحتوى اللفانفي من *E.Coli*، وعدد المكورات المعوية في السمان المعالج بالنبات مقارنة مع السيطرة T0.

الكلمات المفتاحية: الأعشاب الطبية، أداء الجسم، صفات الدم، أنسجة اللفانفي، وزن الجسم.

Received:15/2/2021, Accepted:9/5/2021

INTRODUCTION

Poultry meat is a very important source of protein; therefore, in many countries increased capital has been invested in the poultry industry and especially in quails. As a member of the pheasant family, the quail known as the smallest bird. The amount of protein in the meat is 5-10 percent more than other birds and ruminants (15). Quail meat is also a good source of niacin, phosphorus, copper, thiamine, riboflavin, vitamin B6, zinc, iron, and selenium (32). Nowadays, natural diet additives gained more popularity because of increased health issues regarding synthetic nutrients. Therefore, researcher's persuasion for alternatives to replace synthetic sources (antibiotics and drugs) as diet enhancement in poultry farming becomes an important subject (1). Rifaximin and ranitidine were known as gastrointestinal-selective antibiotics with antiulcer and antimicrobial activities (22, 23). But after the ban of nutritive antibiotics in the European Union, interests in plant additives as poultry feed ingredients become risen continuously (3, 16). Medicinal plants have been known as a source of natural antioxidants, such as phenolic acids, flavonoids, and tannins (12, 36). Numerous research works have shown the efficacy of these natural compounds in stopping the onset and/or progression of many human diseases (17). The curative enhancement efficacy of antioxidants from plants may be related to their role in counteracting reactive oxygen species (ROS) (34). Previous research also suggested the need of adding antioxidants to animal's diets to minimize lipid peroxidation and maximize productivity (8). *Alcea kurdica* (Gule Hero) and *Thymus bovie* (Thyme) are medicinal plants with antioxidant and antimicrobial activities, at which both of them have been used traditionally as anti-inflammatory, anthelmintic, expectorant, antispasmodic, antiseptic agents because of flavonoids, polyphenols, and oxygenated Monoterpenoid contents (2, 31). *Laurus Nobilis* (bay leaf) also known as a traditional medicinal plant with antioxidant activity because of phenolic and flavonoid contents at which its essential oil served as antifungal, the antibacterial activity against various gram positives, negative bacteria, and fungus

species in the various extracts (10). Another study also reported that bay leaf essential oils have been used to increase appetite and enhance immunity (20). In that regard, on that regard, the present study was set to investigate and compare the effects of Hero flower, Thyme and bay leaf extracts in quail drinking water by emphasizing growth performance, intestinal morphology as well as immune, bacterial and physiological status.

MATERIALS AND METHODS

Sample collection and powder preparation

The plant samples were collected in a wide geographic range near Shaqlawa-Erbil/Iraq. Identification and authentication of the plants done by Biology Department/ College of Education/ Salahaddin University-Erbil. The plant part: Hero flowers (*A. kurdica*), Bay leaf (*L. Nobilis*), and Thyme (*T. Bovie*) extracts were prepared as mentioned by Mombeini (25).

Experimental treatments

At age of 1day this study started till 10th weeks, a total of 90 quails was distributed into six treatments, each treatment with 3 replicates and 15 chicks. T0 (control without supplementation). T1 (Rifaximine 200 mg) T2 (Ranitidine 200 mg), T3 (Hero flower extract 100mg), T4 (Thyme extract,100mg), T5 (Bay leaf extract 100mg), each substance was added to a liter of drinking water. All tested groups fed a standard diet contains (2900 ME/kcal, 18.52% crude protein, 4.3% crude fiber, 0.45% methionine, 1.02% lysin), and served ad libitum during the study.

Body performance and histological traits

The birds from each replicate were randomly selected to measuring body weight, and gain, feed intake and Feed Conversion Ratio, water consumption, mortality, and feasibility or economic profit were calculated (30).

Estimation of hematological parameters

At the end of the study 10th weeks blood samples were collected from nine birds by cutting the jugular vein of each group using EDTA tubes for counting total Red Blood Cells and White Blood Cells by using Nutt & Herrick solution, blood collected in capillary tubes to estimate Packed Cell Volume, mean corpuscular volume, mean corpuscular hemoglobin concentration Mean corpuscular hemoglobin (MCH), and Drabkin's Reagent is

used for the quantitative, colorimetric determination of hemoglobin (Hb) concentration in whole blood at 540 nm, heterophil and lymphocyte with Wright-Giemsa stains to determine the heterophils and lymphocytes (H/L) ratio (28).

Serum protein and immunology profile

Serum separated from collected blood and determined total protein, globulin, and albumin, according to the instructions of the kits included in the Buyer's Guide for Life Science Bio-compare. Thus, antibody titers against Gamboro, Newcastle, Infectious bronchitis viral diseases were measured by hemagglutination inhibition assay (HI) test (6).

Ileum bacterial count and histological profile:

At age 10th week, ileum contents were cautiously kept in sterile petri dishes at -20°C until analyses in the laboratory. One gram of each homogenized sample was collected and transferred into 10 ml sterile saline solution for dilution, then each sample was spread on selective agar plates as follows. De Man, Rogosa and Sharpe (MRS) agar medium was used for lactobacillus bacteria and the Nutrient and McConkey agars were utilized for *E. Coli* (19) and Enterococcus were cultured in semisolid Agar and incubated at 37°C for 24 h for the determining colonies. Intestinal ileum samples were dehydrated, cleared, and embedded in paraffin. Serial sections were cut at 5 µm and placed on glass slides. For all assays, sections were deparaffinized in xylene and rehydrated in a graded alcohol series. Sections were examined by light microscopy (35) for determining number of goblet cells, villus height, crypt depth and villus to crypt (V/C) ratio (30).

Statistical analysis

All data were analyzed using CRD (Completely Randomized Design) by the SAS institute program (33). Duncan's multiple range tests were used to compare differences among the treatments.

RESULTS AND DISCUSSION

Medicinal plants as diet additives in poultry farming have been used widely due to multiple biological activities that the herbs that play in stimulate the digestive system and enhancing growth rate (18). The results in Table 1 show that adding medicinal plant extract (Hero flower, bay leaf, and thyme) extracts in the drinking water of quail caused significantly ($P \leq 0.05$) higher body weight and body weight gain in T3, T5, and T4 groups. Data analysis also showed significantly ($P \leq 0.01$) higher economic profit of the marketing age of quails' drunk water mixed plant extracts compared with the other treated groups (T0, T1, and T2). As well as feed conversion ratio (FCR) improved in T5, T3, and T4 in comparison with T2, T1, and T0. While nonsignificant differences among all groups in feed intake. Previous researches reported that Hero flower (9), Thyme (21), and Bay leaf (26) extracts possess great antioxidant activity because of their phenolic, flavonoid, and monoterpenoid contents. A study by Abd El-Galil (1) concluding that medicinal plants with characteristics as an anti-inflammatory, restoratives, antibacterial and anti-parasitic will facilitate the performance productivity in poultry. Studies have shown that expansion of the animal's metabolism by herbal plant powder, generally carbohydrates and proteins in the major nutrient would increase growth rates (31). Another cause of weight gaining of quail treated with plant additives may be because of the plant's aromatic content that stimulating the appetite as known that aroma capable of activating the olfactory nerves and gustatory papillae (5). The results of this study accompanied by previous work on the antioxidant efficacy of plants toward increasing egg and body weight as well as improved Feed Conversion Ratio (FCR) in quails (4). No significant changes have been seen in feed intake of all experimented treatments with no mortality during the experimented period Table 1.

Table 1. Effect of some medicinal plants on body performance of Japanese quail at age 10th wk

Traits	T0	T1	T2	T3	T4	T5	SEM
Initial weight (g)				8.8			
Body weight (g)	197.1 ^b	219.3 ^{ab}	217.0 ^{ab}	246.3 ^a	232.9 ^a	243.7 ^a	12.3
Body weight gain (g)	188.3 ^b	210.0 ^{ab}	208.2 ^{ab}	237.5 ^a	224.1 ^a	234.9 ^a	9.6
Feed intake (g)	725.0 ^a	732.9 ^a	730.8 ^a	729.1 ^a	714.9 ^a	716.4 ^a	8.9
FCR (g feed/ g BWG)	3.85 ^a	3.49 ^{ab}	3.51 ^{ab}	3.07 ^b	3.19 ^b	3.05 ^b	0.38
Economic profit (\$)	0.93 ^c	1.35 ^b	1.39 ^b	1.75 ^a	1.54 ^{ab}	1.70 ^a	0.25

T0: control, T1: Rifaximine 200 mg/L water, T2: Ranitidine 200 mg/L water, T3: Hero flower extract 100 mg/L water, T4: Thyme extract 100 mg/L water, T5: Bay leaf extract 100 mg/L water. ^{a, b, c} means within columns with different superscripts differ significantly at ($P \leq 0.01$) & ($P \leq 0.05$). The same superscripts among the treatments mean insignificant. FCR: Feed conversion ratio

The results in Table 2. show that erythrocytes profile (total counts of RBCs, Hb concentration, and PCV%) and leukocytes profile (total counts of WBCs and lymphocyte%) were significantly ($P \leq 0.01$) increased in the T3 group followed by T4 and T5 experimented treatments. While the lowest erythrocyte and leukocyte profiles were seen in T0, T1, and T2 groups, respectively. The level of MCV, MCH, heterophil%, and H/L ratio were significantly ($P \leq 0.05$) lower in the T3, T5, T4, T2, and T1 treated groups, respectively in comparison with the normal control T0. The level of MCHC was significantly ($P \leq 0.05$) decreased in the T5, T3, T4 groups in comparison with T1 (Rifaximine 2g/L). Positive feedback of Three experimented plants on the erythrocyte and

leukocyte profiles may be linked with their anti-inflammatory and antioxidant activities because of their natural phenolic, flavonoid, and terpenoid contents (9,21, 26). Scientists previously have revealed that adding medicinal plants to poultry drinking water will significantly improve their hematological profile due to the plant's capability in facilitating the erythropoietin process that stimulated kidneys to promote the production of red blood cells and hemoglobin (28). Researchers also have shown the rise of leukocyte production in animals by using medicinal plants and suggest that it could be serve as an immune booster (24). The same results regarding improvement of erythrocyte and leukocyte profiles were found when quails were treated with natural diet additives (27).

Table 2. Effectiveness of some medicinal plants extracts on some hematological parameters of Japanese quail at age 10th wk

Traits	T0	T1	T2	T3	T4	T5	SEM
Total RBCs (10^6 ml^{-1})	3.209 ^c	4.355 ^b	4.310 ^b	4.990 ^a	4.893 ^a	4.886 ^a	0.329
Erythrocytes Profile							
Hb (g/dl)	10.59 ^c	12.35 ^b	12.43 ^b	13.75 ^a	13.79 ^a	13.62 ^a	1.07
PCV (%)	32.79 ^c	37.11 ^b	38.05 ^b	43.29 ^a	43.25 ^a	43.37 ^a	2.16
MCV (fl)	102.18 ^a	85.21 ^b	88.28 ^b	86.75 ^b	89.49 ^b	88.76 ^b	4.68
MCH (Pg)	33.00 ^a	28.35 ^b	28.84 ^b	27.56 ^b	28.18 ^b	27.88 ^b	1.55
MCHC (g/dl)	32.29 ^{ab}	33.28 ^a	32.67 ^{ab}	31.76 ^b	31.88 ^b	31.40 ^b	1.37
Total WBCs (10^3 ml^{-1})	22.45 ^c	27.58 ^b	28.10 ^b	32.06 ^a	31.49 ^a	32.35 ^a	1.83
Leukocytes profile							
Heterophil (%)	40.19 ^a	34.92 ^b	33.18 ^b	20.33 ^d	23.86 ^c	21.09 ^{cd}	1.79
Lymphocyte (%)	59.81 ^c	65.08 ^b	66.82 ^b	79.67 ^a	76.14 ^a	78.91 ^a	2.49
H/L ratio	0.672 ^a	0.537 ^{ab}	0.497 ^b	0.255 ^c	0.313 ^c	0.267 ^c	0.085
Thrombocytes (10^3 ml^{-1})	32.52 ^c	36.95 ^b	38.83 ^b	45.60 ^a	43.97 ^a	45.33 ^a	2.05

T0: control, T1: Rifaximine 200 mg/L water, T2: Ranitidine 200 mg/L water, T3: Hero flower extract 100 mg/L water, T4: Thyme extract 100 mg/L water, T5: Bay leaf extract 100 mg/L water. ^{a, b, c, d} means within columns with different superscripts differ significantly at ($P \leq 0.01$) & ($P \leq 0.05$).

Protein profile (total protein and globulin) concentrations in Table 3 were shown significantly ($P \leq 0.01$) increased amount total protein and globulin in T3, T5, T4, T1, and T2 respectively compared with control T0. However, albumin concentration was

significantly ($P \leq 0.05$) decreased in T3, T5, T4, T1, and T2 treatments, respectively compared with control T0. The same Table, also shows the immune status was represented by antibodies titers against (ND, IBD, and IBV) diseases had significantly ($P \leq 0.05$) increased

levels in the treatments of medicinal plants extract in drinking water than other treated groups. The highest immune titer was recorded in the T3 group against Newcastle (HI), Gambero disease (IBD), and Infectious Bronchitis Viral (IBV) diseases, respectively. Immune statue was found to be almost the same in T1, T4 and T5 experimented groups. The lowest immune titers showed by normal control T0 group against mentioned viral diseases in comparison with other experimented treatments. Past researches have shown the importance of plant extract as feed additives for appetite stimulator, digestive enzyme secretion, and immune enhancements (18). Regarding our data, studies have shown the antioxidants activity of medicinal herbs can significantly delay or prevent lipid peroxidation as well as decrease protein and carbohydrate denaturation (24).The

antioxidant and antimicrobial activities of Hero flower by Bouayed (9), Thyme by Jaradat (21), and Bay leaf by Muñiz-Márquez (26) have been explained previously, at which their phytochemical contents (flavonoids and polyphenols) aid them in scavenging free radicals. Studies also concluded that herbs containing flavonoids, vitamin C, and phenolic compounds will aid in stimulating the immune system by improving lymphocytes, macrophages, and NK cells activities leading to an increase in phagocytosis or interpterion synthesis process (17). The results of this study were similar to the previous study done on the medicinal plant's efficacy on the quail's immunity (27, 37). Similarly, a previous report also showed plant additives caused Physiological and reproductive improvements in Japanese quail (7).

Table 3. Effects of some medicinal plant extract on serum protein profile and immunity by hemagglutination inhibition (HI) test of Japanese quail at age 10th weak.

Traits	T0	T1	T2	T3	T4	T5	SEM
Proteins Profile (g/dl)							
Total protein	2.917 ^c	4.083 ^b	3.982 ^b	4.503 ^a	4.485 ^a	4.488 ^a	0.257
Globulin	1.508 ^c	2.912 ^b	2.773 ^b	3.481 ^a	3.340 ^{ab}	3.413 ^a	0.205
Albumin	1.409 ^a	1.171 ^{ab}	1.209 ^{ab}	1.022 ^b	1.145 ^{ab}	1.075 ^b	0.139
Immune statues (Log 2ⁿ)							
Newcastle disease (HI)	4.50 ^b	7.50 ^a	7.00 ^a	8.00 ^a	7.50 ^a	8.00 ^a	0.495
Gamboro disease (IBD)	3.22 ^b	5.00 ^a	5.33 ^a	5.67 ^a	5.33 ^a	5.33 ^a	0.310
Infectious Bronchitis Viral (IBV)	2.50 ^b	3.67 ^a	3.50 ^a	4.00 ^a	4.00 ^a	3.75 ^a	0.297

T0: control, T1: Rifaximine 200 mg/L water, T2: Ranitidine 200 mg/L water, T3: Hero flower extract 100 mg/L water, T4: Thyme extract 100 mg/L water, T5: Bay leaf extract 100 mg/L water. ^{a, b, c} means within columns with different superscripts differ significantly at ($P \leq 0.01$) & ($P \leq 0.05$).

The results of the ileum bacterial profile in a Table 4 show that the number of *Lactobacillus* counts was significantly ($P \leq 0.01$) higher in T3, T5, and T4, respectively compared with T1, T2, and T0. However, *E. Coli* and *Enterococcus* counts had significantly ($P \leq 0.01$) lower numbers in T3, T4, T5, T2, and T1 compared with the control T0. The results of this study are in accordance with previous findings regarding the antimicrobial activity of Hero flower, which showed great microbial inhibition of aqueous extract of Hero flower against *E. Coli*, *Bacillus*, and some fungal species with no inhibition activity toward *Enterobacter aerogenes* and *Staphylococcus aureus*. Previous works showed the presence of the Phenolic and flavonoid compounds in Hero flower (Gallic acids and quercetin) (9), Thyme (Thymol) (21)

and Bay leaf (kaempferol and quercetin glycosides) (26) could be the reasons behind their inhibitory effect against the growth of *E. coli* and *Enterococcus* bacteria in the ileum of quail's drunk water mixed medicinal plants. Several studies have reported that phenolic compounds mediate the antimicrobial activities of various plant extracts (6,14, 29). The ileum histological study was present in Table 4. The villi height and crypt depth were increased in the plant extract, and antibiotics treated groups. Of which, quails drunk Hero flower extract in the water had significantly ($P \leq 0.01$) higher villus height (865) μm than that (843, 815, 774, 752 and 495) μm for T5, T4, T1, T2, and T0, respectively. The T3 groups also had significantly ($P \leq 0.01$) higher crypt depth (91.92) μm and goblet cells (13.17) numbers than that of T5, T4, T1, T2,

and T0, respectively. Quails with no water treatments (T0) showed poor histological traits regarding villus height (495) μm , crypt depth (80.49) μm , V/C (6.15) μm , and goblet cells number (7.03) μm compared with other treated quails that drunk mixed water additives. Different parts of the small intestine are the areas for absorption of various nutrients, and the structure is related to their function. So, changes in feed and feed additives could be affect the morphology of the intestine at different parts. It is well known that improvement of nutrient absorption occurs by increasing the villi height and decreasing crypt depth. Also, increased intestinal villi height and height to crypt depth ratio and decreased crypt depth by plant extracts could be attributed to the decreased renewal rate of intestinal cells as a result of antioxidant

properties and antimicrobial actions of plant extracts (29). Therefore, Hero flower, Thyme, and Bay leaf extracts could be increasing the absorption of nutrients by affecting the height and crypt depth of the intestine villi and increasing goblet cell numbers. In accordance with this study, supplementation of different levels of thyme oils in quail's diet due to their thymol content (35%) improved intestinal morphology (11). In contrast, the results of a study on rats showed that the villi width significantly increased in the duodenum and jejunum of the rats fed thyme volatile oil (23). Accordingly, previous reports showed a significantly increased villi height to crypt depth ratio by thymol compared to the basal diet in a state where birds are exposed to pathogens (13).

Table 4. Effect of some medicinal plants extracts on ileum bacterial and histological traits of Japanese quail at age 10th wk

Traits		T0	T1	T2	T3	T4	T5	SEM
Bacterial profile (CFU \times 105/g)	<i>E. Coli</i>	8.02 ^a	3.01 ^b	3.15 ^b	3.11 ^b	3.00 ^b	3.00 ^b	0.428
	<i>Enterococcus</i>	6.83 ^a	2.55 ^b	2.51 ^b	2.55 ^b	2.88 ^b	2.68 ^b	0.503
	<i>Lactobacillus</i>	4.92 ^c	12.08 ^b	12.08 ^b	13.08 ^a	12.70 ^a	13.00 ^a	0.719
Ileum histology	Villus height (μm)	495 ^c	774 ^b	752 ^b	865 ^a	815 ^{ab}	843 ^a	76.0
	Crypt depth (μm)	80.49 ^c	87.26 ^{ab}	84.97 ^b	91.92 ^a	87.92 ^{ab}	89.87 ^{ab}	2.79
	V/C	6.15 ^b	8.87 ^a	8.85 ^a	9.41 ^a	9.27 ^a	9.38 ^a	0.920
	Goblet cells No. ¹	7.03 ^c	11.69 ^b	11.66 ^b	13.17 ^a	12.66 ^{ab}	12.94 ^a	1.02

T0: control, T1: Rifaximine 200 mg/L water, T2: Ranitidine 200 mg/L water, T3: Hero flower extract 100 mg/L water, T4: Thyme extract 100 mg/L water, T5: Bay leaf extract 100 mg/L water. V/C: height to crypt depth ratio. ^{a, b, c} means within columns with different superscripts differ significantly at ($P \leq 0.01$) & ($P \leq 0.05$).

CONCLUSION

The current research draws attention to compare three traditional medicinal plant additives in quail's drinking water with two antibiotic drugs and a control group. Their effect caused an increase in the body weight and gain, economic profit (feasibility) of birds by maximizing the absorbed nutrients based on their positive effect on the hematology, immune status, intestinal morphology, and *Lactobacillus* population in the ileum of quails. *A.kurdica*, *L.nobilis*, and *T.bovie* extracts were more influential due to their higher contents of gallic acid, quercetin, and kaempferol glycosides compared with the antibiotic drugs and control groups. Moreover, *A.kurdica* extract could compete with the antibiotics to fight intestinal microbes such *E.Coli* and *Enterococcus*. As result, adding plant extracts to quails drinking water could be a better choice for increasing poultry production than antibiotics with fewer adverse

effects, which positively reflects on the profitability of birds.

REFERENCES

1. Abd El-Galil, K. and Henda AM. 2015. Effect of ginger roots meal as feed additives in laying Japanese quail diets. *The Journal of American Science*, 11(2):164-173.
2. Aboelsoud, N. H. 2010. Herbal medicine in ancient Egypt. *Journal of Medicinal Plants Research*, 4(2), 082–086.
3. Abouelezz, FMK. 2017. Evaluation of *Spirulina* Algae (*Spirulina platensis*) as a feed supplement for Japanese quail: nutritional effects on growth performance, egg production, egg quality, blood metabolites, sperm-egg penetration and fertility. *Egyptian Poultry Science Journal*, 37(3):707–719
4. Abudoulrahman, K.K., M.A Mustafa, and A.A., Abduljabbar. 2019. The effect of heat stress on oxidative stress and antioxidant status in local quail hens supplemented with onion

- and garlic oils. Tikrit Journal for Agricultural Sciences, 19(1):103-110
5. Alagawany, M., S.S Elnesr, and M.R Farag. 2019. Use of liquorice (*Glycyrrhiza glabra*) in poultry nutrition: Global impacts on performance, carcass and meat quality. In World's Poultry Science Journal. 75(2), 293-304
 6. Alshawsh, M.A., M.A. Abdulla, S. Ismail, Z.A. Amin, S.W. Qader, H.A. Hadi, and N.S. Harmal. 2012. Free radical scavenging, antimicrobial and immunomodulatory activities of orthosiphon stamineus. Molecules, 17(5): 5385–5395.
 7. Al salhie K., F.S. Shawkat and B. Lahmood. 2017. Effect of supplementation date palm pollen on some physiological and reproductive traits of Japanese quail birds (*Coturnix coturnix japonica*). Iraqi Journal of Agricultural sciences, 48(6):1389-1398
 8. Aminzade, B., B. Karami and E. Lotfi. 2012. Meat quality characteristics in Japanese quails fed with *Mentha piperita* plant. Animal biology and animal husbandry International Journal of the Bioflux Society, 4(1), 20–23.
 9. Bouayed, J., Kh. Piri, H. Rammal, A. Dicko, F. Desor, Ch. Younos and R. Soulimani. 2007. Comparative evaluation of the antioxidant potential of some Iranian medicinal plants. 104 (2007) 364–368.
 10. De Corato, U., O. Maccioni, M. Trupo and G. Di Sanzo. 2010. Use of essential oil of *Laurus nobilis* obtained by means of a supercritical carbon dioxide technique against postharvest spoilage fungi. Crop Protection, 29(2), 142-147
 11. Dehghani, N., M. Afsharmanesh, M. Salarmoini, H. Ebrahimnejad and A. Bitaraf. 2018. Effect of pennyroyal, savory and thyme essential oils on Japanese quail physiology. Heliyon, 4(10), e00881.
 12. Djeridane, A., M. Yousfi, B. Nadjemi, D. Boutassouna, P. Stocker and N. Vidal. 2006. Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. Food Chemistry, 97: 654–660
 13. Du, E., W. Wang, L. Gan, Z. Li, S. Guo and Y. Guo. 2016. Effects of thymol and carvacrol supplementation on intestinal integrity and immune responses of broiler chickens challenged with *Clostridium perfringens*. Journal of Animal Science and Biotechnology, 7(1), 19–29.
 14. Ebrahimabadi, A. H., A. Mazoochi, F.J. Kashi, Z. Djafari-Bidgoli and H. Batooli. 2010. Essential oil composition and antioxidant and antimicrobial properties of the aerial parts of *Salvia eremophila* boiss. from Iran. Food and Chemical Toxicology, 48(5), 1371–1376.
 15. Genchev, A., G. Mihaylova, S. Ribarski, A. Pavlov and M. Kabakchiev. 2008. Meat quality and composition in Japanese quails. Trakia Journal of Sciences, 6(4), 72–82
 16. Gerzilov, V., A. Nikolov, P. Petrov, N. Bozakova, G. Penchev and A. Bochukov. 2015. Effect of a dietary herbal mixture supplement on the growth performance, egg production and health status in chickens. Journal of Central European Agriculture, 16(2): 10-27.
 17. Halliwell, B., J.M.C. Gutteridge and C.E. Cross. 1992. Free radicals, antioxidants, and human disease: where are we now? Journal of Laboratory and Clinical Medicine, 119, 598–619.
 18. Hamodi, S.J. and F.M. Al-Khilani. 2014. The effect of supplemental different level of roselle flower in diet on Japanese quail performance. Scientific Papers. Series D. Animal Science, LVII:94–98
 19. Harrigan, W.F., and M.F. McCance. 1976. Laboratory methods. in: Food and Dairy Microbiology Academic Press .London.
 20. Jamroz, D., A. Wiliczkievicz, T. Wertelecki, J. Orda and J. Skorupińska. 2005. Use of active substances of plant origin in chicken diets based on maize and locally grown cereals. British Poultry Sci, 46(4): 485–493
 21. Jaradat, N., L. Adwan, S. K'aibni, N. Shraim and A.N. Zaid. 2016. Chemical composition, anthelmintic, antibacterial and antioxidant effects of thymus bovei essential oil. BMC Complementary Alternative Medecine, 16: 418.
 22. Konorev, M.R. 1998. Effect of ranitidine and metronidazole on IgG activity of patients with *Helicobacter pylori* infection. International Journal of Antimicrobial Agents, 10(3), 249–250.
 23. Koo, H.L., and H.L. DuPont. 2010. Rifaximin: a unique gastrointestinal-selective

antibiotic for enteric diseases. *Current Opinion in Gastroenterology*, 26(1), 17–25.

24. Ladokun, O., M. Ojezele and O. Arojojoye. 2015. Comparative study on the effects of aqueous extracts of *viscum album* (*Mistletoe*) from three host plants on hematological parameters in albino rats. *African Health Sciences*, 15(2):606-612.
25. Mombeini, T., H. Gholami Pourbadie, M. Kamalinejad, S. Mazloumi and A.R. Dehpour. 2017. Anxiolytic-like and sedative effects of *Alcea Aucheri* (*Boiss.*) Alef. flower extract in the laboratory rat. *Iran Journal Pharmacy Research*; 16(4): 1495–1508.
26. Muñiz-Márquez, D.B., G.C. Martínez-Ávila, J.E. Wong-Paz, R. Belmares-Cerda, R. Rodríguez-Herrera and C.N. Aguilar. 2013. Ultrasound-assisted extraction of phenolic compounds from *Laurus nobilis* L. and their antioxidant activity. *Ultrasonics Sonochemistry*. 20 (5):1149-1154
27. Mustafa, M.A., P.S. Sabir and N.A. Mustafa. 2017. Effect of functional feed additives on egg production, hatchability and hematological traits of Japanese quails during summer condition. *Iraqi Journal of Agricultural Sciences*. 48(special Issue): 80–85.
28. Oleforuh-Okoleh, V. U., S.O. Olorunleke, and I.J. Nte. 2015. Comparative response of bitter leaf (*Vernonia amygdalina*) infusion administration on performance, haematology and serum biochemistry of broiler chicks. *Asian Journal of Animal Sciences*, 9(5):217-224
29. Patra, A. K., S. Amasheh and J.R. Aschenbach. 2018. Modulation of gastrointestinal barrier and nutrient transport function in farm animals by natural plant bioactive compounds – a comprehensive review. *Critical Reviews in Food Science and Nutrition*, 59(20), 3237–3266.
30. Pooryousef, M.M and M.N. Hosseini. 2012. Comparative effect of Liquorice root extract medicinal plants and probiotic in diets on performance, carcass traits and serum composition of Japanese quails. *Global Veterinarian*, 8(1): 39-42.
31. Qader, S.W., and H.M. Awad. 2014. Evaluation of antioxidant, antimicrobial and cytotoxicity of *Alcea kurdica* Alef. *Jordan Journal of Biological Sciences*. 7(3): 205–209.
32. Santhi, D., and A. Kalaikannan. 2017. Japanese quail (*Coturnix coturnix japonica*) meat: characteristics and value addition. *World's Poultry Science Journal*, 73 (2): 337-344
33. SAS, Statistical Analysis System. 2005. User's Guide for Personal Computer. Release 8.2 SAS Institute Inc. Cary. NC, USA
34. Subedi, L., S. Timalsena, P. Duwadi, R. Thapa, A. Paudel and K. Parajuli. 2014. Antioxidant activity and phenol and flavonoid contents of eight medicinal plants from Western Nepal. *Journal of Traditional Chinese Medicine*, 34 (5): 584-590
35. Uni, Z., S. Ganot and D. Sklan. 1998. Posthatch development of mucosal function in the broiler small intestine. *Poultry Science*, 77(1):75-82
36. Wong, C., H. Li, K. Cheng and F. Chen. 2006. A systematic survey of antioxidant activity of 30 Chinese medicinal plants using the ferric reducing antioxidant power assay. *Food Chemistry*, 97(4):705–711
37. Mustafa, M.A. and P.H. Wasman. 2020. The impact of powders and oil additives of cinnamon and clove in quail diet as antistressor and antioxidant during hot months. *Iraqi Journal of Agricultural Science*, 51(3),760-766.